

Mouse IL-10 ELISA Kit

Vertrieb:

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Hinweis/Note:

Der Packungsbeileger dient nur als erste Information. Der relevante Packungsbeileger liegt der Ware bei.

The datasheet is only a first information.

The relevant datasheet is included with the product.

For any questions regarding troubleshooting or performing the assay, please contact our support team at support@assaypro.com.

Thank you for choosing Assaypro.

Assay Summary

Add 50 μl of Standard/ Sample per well. Incubate 2 hours.



Wash, then add 50 µl of Biotinylated Antibody per well. Incubate 2 hours.



Wash, then add 50 µl of SP Conjugate per well. Incubate 30 minutes.



Wash, then add 50 µl of Chromogen Substrate per well. Incubate 20 minutes.



Add 50 μ l of Stop Solution per well. Read at 450 nm immediately.

Assay Template

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AssayMax Mouse Interleukin-10 (IL-10) ELISA Kit

Catalog No. EMI3010-1 Sample Insert/Reference Only

Introduction

Interleukin-10 (IL-10) is a regulatory cytokine, and its principal role in vivo is to limit inflammatory response. IL-10 has been shown to influence both the susceptibility and course of various diseases (1). Interleukin-10 (IL-10) is a key cytokine produced by a multitude of immune effector cells and possesses distinct regulatory effects on immune functioning in the skin (2). The accelerated alveolar bone loss observed in IL-10 (-/-) mice is a late-onset condition and that lack of IL-10 may have an effect on bone homeostasis (3).

Principle of the Assay

The AssayMax Mouse IL-10 ELISA (Enzyme-linked Immunosorbent Assay) kit is designed for detection of mouse IL-10 in mouse plasma, serum, tissue extracts, and cell culture samples. This assay employs a quantitative sandwich enzyme immunoassay technique that measures mouse IL-10 in less than 5 hours. A polyclonal antibody specific for mouse IL-10 has been pre-coated onto a 96-well microplate with removable strips. IL-10 in standards and samples is sandwiched by the immobilized antibody and a biotinylated polyclonal antibody specific for mouse IL-10, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is then washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

Caution and Warning

- Prepare all reagents (working diluent buffer, wash buffer, standards, biotinylated antibody, and SP conjugate) as instructed, prior to running the assay.
- Prepare all samples prior to running the assay. The dilution factors for the samples are suggested in this protocol. However, the user should determine the optimal dilution factor.
- Spin down the SP conjugate vial and the biotinylated antibody vial before opening and using contents.
- This kit is for research use only.
- The kit should not be used beyond the expiration date.

• The Stop Solution is an acidic solution.

Reagents

- **Mouse IL-10 Microplate:** A 96 well polystyrene microplate (12 strips of 8 wells) coated with a polyclonal antibody against mouse IL-10.
- **Sealing Tapes:** Each kit contains 3 precut, pressure sensitive sealing tapes that can be cut to fit the format of the individual assay.
- Mouse IL-10 Standard: Mouse IL-10 in a buffered protein base (2 ng, lyophilized).
- **Biotinylated Mouse IL-10 Antibody (100x):** A 100-fold concentrated biotinylated polyclonal antibody against mouse IL-10 (80 μl).
- MIX Diluent Concentrate (10x): A 10-fold concentrated buffered protein base (30 ml).
- Wash Buffer Concentrate (20x): A 20-fold concentrated buffered surfactant (30 ml, 2 bottles).
- Streptavidin-Peroxidase Conjugate (SP Conjugate): A 100-fold concentrate (80 µl).
- **Chromogen Substrate**: A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- **Stop Solution**: A 0.5 N hydrochloric acid to stop the chromogen substrate reaction (12 ml).

Storage Condition

- Upon arrival, immediately store components of the kit at recommended temperatures up to the expiration date.
- Store SP Conjugate and Biotinylated Antibody at -20°C.
- Store Microplate, Diluent Concentrate (10x), Wash Buffer, Stop Solution, and Chromogen Substrate at 2-8°C.
- Unused microplate wells may be returned to the foil pouch with the desiccant packs and resealed. May be stored for up to 30 days in a vacuum desiccator.
- Diluent (1x) may be stored for up to 30 days at 2-8°C.
- Store Standard at 2-8°C before reconstituting with diluent and at -20°C after reconstituting with diluent.

Other Supplies Required

- Microplate reader capable of measuring absorbance at 450 nm.
- Pipettes (1-20 μl, 20-200 μl, 200-1000 μl, and multiple channel).
- Deionized or distilled reagent grade water.

Sample Collection, Preparation and Storage

- Plasma: Collect plasma using one-tenth volume of 0.1 M sodium citrate as an anticoagulant. Centrifuge samples at 3000 x g for 10 minutes and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles (EDTA or Heparin can also be used as an anticoagulant).
- **Serum:** Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 3000 x g for 10 minutes. Remove serum and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **Cell Culture Supernatants:** Centrifuge cell culture media at 3000 x g for 10 minutes to remove debris. Collect supernatants and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **Tissue:** Extract tissue samples with 0.1 M Tris-buffered saline (pH7.4) containing 0.5% Triton X-100 and centrifuge at 14000 x g for 30 minutes. Collect the supernatant and measure the protein concentration. Dilute the tissue extract if necessary and assay. Freeze the remaining extract at -20°C or below. Avoid repeated freeze-thaw cycles.

Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use.
- MIX Diluent Concentrate (10x): If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved. Dilute the MIX Diluent Concentrate 1:10 with reagent grade water. Store for up to 30 days at 2-8°C.
- Standard Curve: Reconstitute the 2 ng of Mouse IL-10 Standard with 1 ml of MIX Diluent to generate a 2 ng/ml standard solution. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare duplicate or triplicate standard points by serially diluting the standard solution 1:2 with equal volume of MIX Diluent to produce 1, 0.5, 0.25, 0.125, 0.063, and 0.031 ng/ml solutions. MIX Diluent serves as the zero standard (0 ng/ml). Any remaining solution should be frozen at -20°C and used within 30 days.

Standard Point	Dilution	[Mouse IL-10] (ng/ml)
P1	Standard (2 ng/ml)	2.000
P2	1 part P1 + 1 part MIX Diluent	1.000
Р3	1 part P2 + 1 part MIX Diluent	0.500
P4	1 part P3 + 1 part MIX Diluent	0.250
P5	1 part P4 + 1 part MIX Diluent	0.125
P6	1 part P5 + 1 part MIX Diluent	0.063
P7	1 part P6 + 1 part MIX Diluent	0.031
P8	MIX Diluent	0.000

- **Biotinylated Mouse IL-10 Antibody (100x):** Spin down the antibody briefly and dilute the desired amount of the antibody 1:100 with MIX Diluent. Any remaining solution should be frozen at -20°C.
- Wash Buffer Concentrate (20x): If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved. Dilute the Wash Buffer Concentrate 1:20 with reagent grade water.
- **SP Conjugate (100x):** Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with MIX Diluent. Any remaining solution should be frozen at -20°C.

Assay Procedure

- Prepare all reagents, standard solutions, and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-25°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccants inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.
- Add 50 μ l of Mouse IL-10 Standard or sample per well. Cover wells with a sealing tape and incubate for 2 hours. Start the timer after the last addition.
- Wash five times with 200 μ l of Wash Buffer manually. Invert the plate each time and decant the contents; hit 4-5 times on absorbent material to completely remove the liquid. If using a machine, wash six times with 300 μ l of Wash Buffer and then invert the plate, decanting the contents; hit 4-5 times on absorbent material to completely remove the liquid.
- Add 50 μ l of Biotinylated Mouse IL-10 Antibody to each well and incubate for 2 hours.
- Wash the microplate as described above.
- Add 50 μ l of Streptavidin-Peroxidase Conjugate per well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash the microplate as described above.

- Add 50 μ l of Chromogen Substrate per well and incubate for approximately 20 minutes or till the optimal blue color density develops. Gently tap the plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 μ l of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm immediately. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections. Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

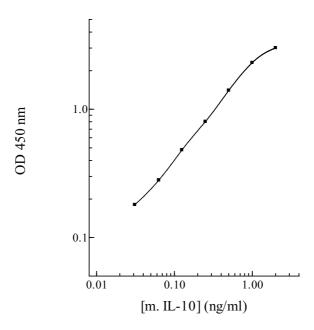
Data Analysis

- Calculate the mean value of the triplicate readings for each standard and sample.
- To generate a standard curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit.
- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

Standard Curve

• The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.

M. IL-10 Standard Curve



Performance Characteristics

- The minimum detectable dose of mouse IL-10 is typically ~ 0.03 ng/ml.
- Intra-assay and inter-assay coefficients of variation were 4.9% and 7.1% respectively.
- This assay recognizes both natural and recombinant mouse IL-10.

Linearity

	Average Percentage of Expected Value		
Sample Dilution	Plasma	Serum	
No Dilution	95%	97%	
1:2	100%	101%	
1:4	98%	100%	

Recovery

Standard Added Value	0.0625 – 1 ng/ml
Recovery %	82 – 115%
Average Recovery %	99%

Cross-Reactivity

Species	% Cross Reactivity
Canine	None
Bovine	None
Monkey	None
Human	None
Rat	None
Swine	None
Rabbit	None

References

- (1) Opdal SH (2004) FEMS Immunol Med Microbiol. 42(1): 48-52
- (2) Weiss E et. al. (2004) J Am Acad Dermatol. 50(5): 657-75; quiz 676-8.
- (3) Al-Rasheed A et. al. (2004) J Periodontal Res. 39(3): 194-8

Version 1.2R

Related products

- El3010-1 AssayMax Human IL-10 ELISA Kit (Plasma, Serum, Cell Culture, and Tissue samples)
- ERI3010-1 AssayMax Rat IL-10 ELISA Kit (Cell Culture samples)

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